

Production and Evaluation of Papain and Pectinesterase Enzymes from Papaya Fruits

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Abstract

The objective of this study was to produce and evaluate the papain and Pectinesterase enzymes from papaya (*Carica papaya L.*) fruit. The protein content and proteolytic activity were found to be 0.74mg/ml and 7.68 U/ml of papain enzyme, 0.48mg/ml and 2.8 U/ml of papaya pectinesterase, respectively. The optimum NaCl concentration on PE activity was found to be 0.3M. The optimum pH on the reaction activity of crude papain and papaya pectinesterase were found to be 6.4 and 7.0, respectively. The optimum temperature was found to be 60°C and 55°C, respectively. The optimum enzyme concentration was found to be 0.6g/100ml and 12ml of the extract, respectively. The optimum substrate concentration was found to be 1.1g/100ml and 6.5 mg/ml. The Michaelis–Menten constant and maximal rate of the crude papain and papaya Pectinesterase equaled to 0.43 g/100ml potassium phosphate buffer solution and 13.68 U/ml for the first enzyme while, these values were 3.2 mg/ml and 15.6 U/ml for the second ones, respectively.

Keywords: Papain, Pectinesterase, Papaya fruits, Proteolytic activity, Kinetic parameters.

Introduction

Papaya fruit (*Carica papaya L.*) or the pawpaw is a popular and economically important fruit tree of tropical and subtropical countries. The fruit is consumed world-wide as fresh fruit and as a vegetable or used as processed products (Teixeira da Silva *et al.*, 2007). Its high content of vitamin C, E and provitamin A, which has a protective effect against cancers, so adding a daily serving of papaya to diets may lessen the risk of developing cancer. Also, rich in fiber, which travels through the body and binds itself to cancer-causing toxins in the colon and its low calorie status that is recommended for low hypo-caloric diets. Pamela and James (2010).

The papain (E.C. 3.4.22.2.) is a natural proteolytic enzyme that was extracted from the latex of papaya's unripe fruits (Baeza *et al.*, 1989) that digest protein and used as meat tenderizer, as digestive medicine, in pharmaceutical, brewing and tanning industries, and in manufacturing of chewing gum, Puiget *et al.* (2008).

Rao and Rangan (2001) studied the production of crude papain from papaya latex and by-product. Latex containing papain was extracted from 70- to 100-day-old fruits of pawpaw by tapping 4 or 6 times on the fruit. Latex was subsequently collected from the same fruit at 4- and 7-day intervals. Dayanand (2013) studied the evaluation of comparative total proteolytic activity in plant lattices. He found that the total proteolytic activity of *Carica papaya* latex (crude papain) solubilized in water at pH 7.6 and 37°C was 1.750 units, 128 mg total protein, 13.6 (activity/mg protein) specific activity. Also, he evaluated the total proteolytic activity at different pH conditions and the specific activity by using protein as substrate, respectively. Pectinesterase (PE, pectin methylesterase, pectin pectylhydrolase, EC, 3.1.1.11) removes methoxyl groups from methylated galacturonic residues of pectic substances, (Vanden

broecketal., 1999). This enzyme is widely distributed in higher plants and can be found in different plant tissues; mainly those contained in fruits (Baron and Thibault, 1985). Pectinesterase play a central role in the process of fruit softening during ripening, in addition influences the quality and stability of processed products. A short period after papaya is pulped into puree, a gel is formed. This gel formation has been attributed to the enzymatic action of pectinesterase. Also, of great concern to the citrus industry since it has been definitely established that it is the causative agent for clarification of citrus juices and gelation of concentrates, Sarr and Tsai (2008), the enzyme must be inhibited by pasteurization at 90 °C for one minute.

The aim of this investigation is to study the production and evaluation of papain and Pectinesterase enzymes from papaya fruits and kinetic parameters of these enzymes.

Materials and Methods

Papaya (*Carica papaya L.*) fruits solo variety was obtained from farm agricultural research station, Faculty of agriculture at Moshtohor, Benha University. Samples were collected in 2012 and 2013.

Preparation of papaya fruit pulp:-

The fruits were used at two maturity stages, mature green and ripe stages (100% yellow). Ripe fruits were washed and left to drain. Then manually were peeled, deseeded and then were cut to small pieces and converted into smooth papaya pulp using mixer or blender (model–brown). Samples from the fresh pulp were used to the evaluation of pectinesterase (PE). The remaining pulp was stored in containers at the deep freezer for further processing. Papaya latex used in this work was collected from unripe papaya

fruits to study the kinetics parameters of produced papain enzyme.

Extraction of crude papain (papaya latex protease) and papaya pectinesterase enzyme (PE).

The extraction of papaya latex protease was carried out according to the method described by (Nitsawang, *et al.* 2006). Also, the extraction of papaya Pectinesterase was performed according to the method described by Wicker (1992).

Effect of NaCl on PE extraction and activity from papaya fruits:-

The effect of NaCl concentrations (0.5 – 3.0 M) on the extraction of papaya pectinesterase was studied. Also, the effect of NaCl concentration on PE activities was tested in the range (0.05 – 0.5 M) NaCl in the reaction mixture. Measurements were carried out by using 1% pectin dissolved in NaCl solution of varying concentrations at pH 7.0 and at 30°C according to the method of Wicker (1992).

Proteolytic activity and pectinesterase activity:-

Proteolytic activity was determined according to the method described by Arnon, (1970). Pectinesterase (PE) activity was assayed by the titration method proposed by Kertesz (1955). Protein enzyme content was determined according to the method described by Bradford (1976), using bovine serum albumin (BSA) as standard.

Kinetic parameters of crude papain and papaya pectinesterase enzymes:-

The effect of pH on the crude papain enzyme activity was measured at different pH values ranging from (5.8 – 8.0) by using 0.05 M phosphate buffer. Also, the effect of pH on the papaya pectinesterase enzyme activity was determined at different pH values ranging from (5.0- 9.0) by using 10 ml of 1% pectin solution containing 0.15 M NaCl solution. All reactions were carried out for 10 min at different pH values and at 30°C. The activities of crude papain and papaya pectinesterase were measured at different degrees of temperatures ranging from 30 – 80°C. The effect of enzyme concentration on crude papain activity was measured at different enzyme concentrations values ranging from (0.1- 1 %). However, the activity of papaya (PE) was measured by using different volumes of enzyme extract solution. A valid assay was established by using different enzyme concentrations (1-10 ml) and 10 ml of 1% pectin in 0.15M NaCl solution, at optimum pH and temperature. Solutions of casein varying in concentrations (0.1, 0.2, 0.3, 0.4, 0.5, 0.6, 0.7, 0.8, 0.9, 1.0, 1.1, 1.2, 1.3, 1.4 and 1.5%, w/v) were employed to study the effect of substrate concentration on crude papain activity at optimum pH, temperature and constant amount of enzyme concentration according to the methods of (Arnon, 1970). Also, solutions of pectin varying in

concentrations (0.5, 1.0, 1.5, 2, 2.5, 3, 3.5, 4, 4.5, 5, 5.5, 6, 6.5, 7, 7.5, 8, 8.5, 9, 9.5 and 10 mg/ml) were employed to study the effect of substrate concentration on pectinesterase activity at optimum pH, temperature and constant amount of pectinesterase enzyme according to the methods of Fayyaz *et al.* (1995). Michaelis constant (K_m) and maximum reaction velocity (V_{max}) were calculated, then Lineweaver and Burk (1954) plots were determined.

Results and Discussion

Extraction of papain and papaya pectinesterase from papaya fruits:-

The amount of fresh latex obtained by the first fruit tapping was 6.0 g/kg fruit, and after the drying process the amount of crude papain obtained was 1.52 g/kg fruit, the moisture content was found to be 74.6% of papaya latex protease. Also, the effect of sodium chloride concentrations (0.5- 3 M) on the extraction of papaya pectinesterase (PE) is presented in Fig (1). Raising the NaCl concentration in the extracting solution from (0.5 to 1.0 M) increases the activity of PE, reaching its maximum activity (6.25 units /mg protein) at 1.5 M of NaCl. This concentration of NaCl for extracting PE for papaya fruits was similar to value obtained by Fayyaz and Asbi (1993), they found 1.7M the optimum concentration of PE extracting from guava, and also it is lower than reported by Maria *et al.* (2007), who found that 2 M is the optimum NaCl concentration for hawthorn PE extraction. The strong influence of NaCl on papaya PE extraction suggested that the enzyme associated with the cell wall by ionic interaction. The decrease in PE activity with NaCl concentrations higher than 1.5 M might arise as an effect of salting out (Polacsek-Ra'cz and Posar-Hajnal, 1976). These variations between the obtained and other results may be due to the different sources produced these enzymes.

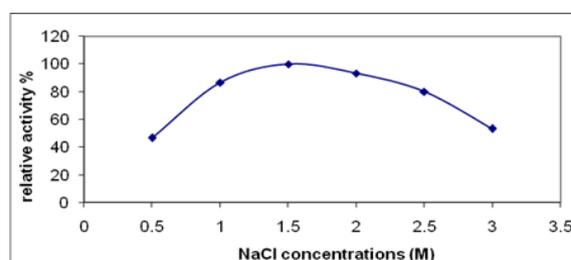


Fig. (1). Effect of NaCl concentrations on papaya pectinesterase extraction.

Factors affecting on the reaction velocity and activity of crude papain and pectinesterase enzymes:-

The proteolytic activity and protein content of crude papain enzyme was determined and the obtained results are illustrated in Table (1). From these data

the activity was found to be 7.68 U/ml, 0.74 mg/ml of protein content and 10.38 U/mg protein of specific activity. The obtained results are in agreement with that stated by **Rofael and Salwa (2003)**. On the other hand, the activity and protein content of pectinesterase enzyme was determined and the

results are tabulated in Table (1). From these data the activity was found to be 2.8 micro-equivalent COOH/min/ml crude extract, 0.48 mg/ml of protein content and 5.83 units/mg protein of specific activity the obtained results are in agreement with that stated by **Fayyazet al. (1995)**.

Table 1. Activity and protein content of crude papain and papaya Pectinesterase enzymes.

Extracted Enzymes	Obtained of tyrosine equivalents (µm/ml)	Activity (U/ml)	Protein content (mg/ml)	Specific activity (U/mg protein)
Crude papain	0.64	7.68	0.74	10.38
papaya pectinesterase	-----	2.8	0.48	5.83

Effect of NaCl concentrations on the reaction activity of papaya pectinesterase (PE) enzyme:-

The effect of NaCl concentration on PE activity was tested in the range (0.05- 0.5 M) NaCl in the reaction mixture. Measurements were carried by using 1% pectin, pH 7.0 at 30°C.

The accomplished results are shown in Table(2) and Fig (2). The obtained result indicates that papaya pectinesterase activity like other plant pectinesterase

also depended on the NaCl concentration in the assay. The effectiveness of the salt was related to its concentration in the assay mixture and the activity increased with the concentration up to 0.3 M, but on raising NaCl level further the activity decreased. This optimum level of NaCl concentration obtained in the present study is lower than that observed by **Maria et al. (2007)** and is higher to the value of (0.25M) NaCl reported by **Fayyazet al. (1995)**.

Table 2. Effect of NaCl concentration on the reaction activity of papaya pectinesterase (PE) enzyme.

NaCl Conc. (M)	Activity (units/ml)	Specific activity U/mg protein	Relative activity (%)
0.05	3.6	7.50	23.07
0.10	6.6	13.75	42.30
0.15	8.4	17.50	53.85
0.20	11.4	23.75	73.07
0.25	13.8	28.75	88.46
0.30	15.6	32.50	100.00
0.35	15.0	31.25	96.15
0.40	12.0	25.00	76.92
0.45	7.8	16.25	50.00
0.50	7.2	15.00	46.15

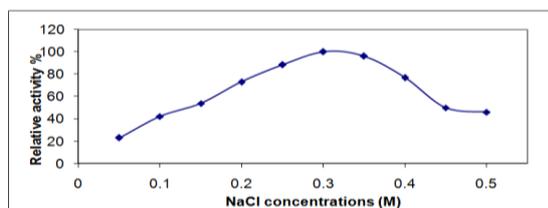


Fig (2). Effect of NaCl concentration on the reaction activity of papaya pectinesterase (PE) enzyme.

Effect of pH on the reaction activity of crude papain and papaya pectinesterase (PE) enzymes:-

The effect of pH on the reaction activity of crude papain was evaluated at different pH values. The reaction mixtures were prepared in different solutions at pH values of 5.8, 6.0, 6.2, 6.4, 6.6, 6.8, 7.0, 7.2, 7.4, 7.6, 7.8 and 8.0 with 0.65% (w/v) casein by using potassium phosphate buffer at pH 7.5 and 37°C. The enzyme activities obtained are presented in

Table (3) and Fig. (3) The maximum enzyme activity i.e. 8.16units/ml was observed at pH 6.4 for soluble crude papain enzyme. The bell-shaped curve (Fig., 3) was obtained with a relatively plateaus and with sharply decreasing rate on the right side. The present papain pH optimum of pH 6.4 is lower than the pH optima of (7.0) reported for the optimum conditions for the activities of both crude extracted enzyme and commercial papain, **Pitprecha and Damrongsakkul (2006)**.

On the other hand, the obtained results for papaya pectinesterase activity as a function of pH is shown in Table (4) and illustrated in Fig (4) The obtained results, indicated that the activity gave pH optima of 7.0 The present PE pH optima of 7.0 was slightly higher than the pH optima of 6.5 reported by pectinesterase from plant tomato, **Nikolic and Mojovic (2007)**. This value differed from that

reported by Denes *et al.* (2000) and Fayyazet *al.* (1995).

Table 3. Effect of pH on the reaction activity of crude papain enzyme.

pH's	Obtained of tyrosine equivalents ($\mu\text{m}/\text{ml}$)	Activity (units/ml)	Specific activity U/mg	Relative activity (%)
5.8	0.62	7.44	10.05	91.17
6	0.63	7.56	10.21	92.64
6.2	0.64	7.68	10.37	94.11
6.4	0.68	8.16	11.03	100.00
6.6	0.62	7.44	10.05	91.17
6.8	0.61	7.32	9.89	89.70
7.2	0.60	7.2	9.72	88.23
7.4	0.58	6.96	9.40	85.29

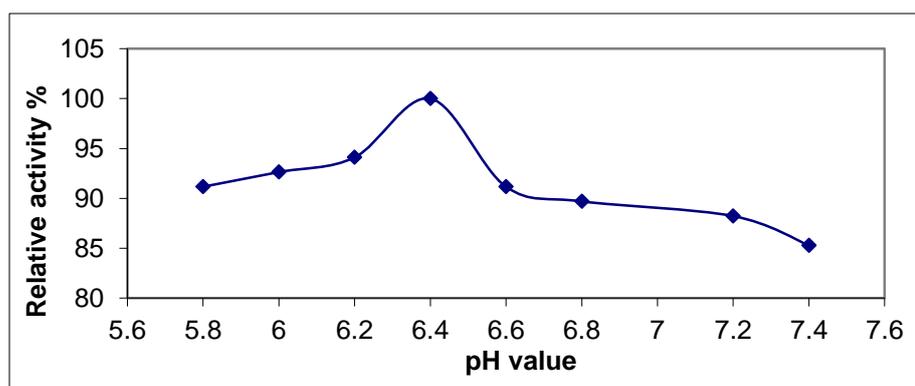


Fig (3): Effect of pH on the reaction activity of crude papain enzyme.

Table 4. Effect of pH on the reaction activity of crude papaya pectinesterase (PE).

pH's	Activity (units/ml)	Specific activity U/mg protein	Relative activity (%)
5.0	5.4	11.25	31.03
5.5	7.8	16.25	44.83
6.0	10.8	22.5	62.06
6.5	14.4	30.0	82.76
7.0	17.4	36.25	100.00
7.5	16.2	33.75	93.10
8.0	14.4	30.0	82.75
8.5	10.8	22.5	62.07
9.0	6.0	12.5	34.48

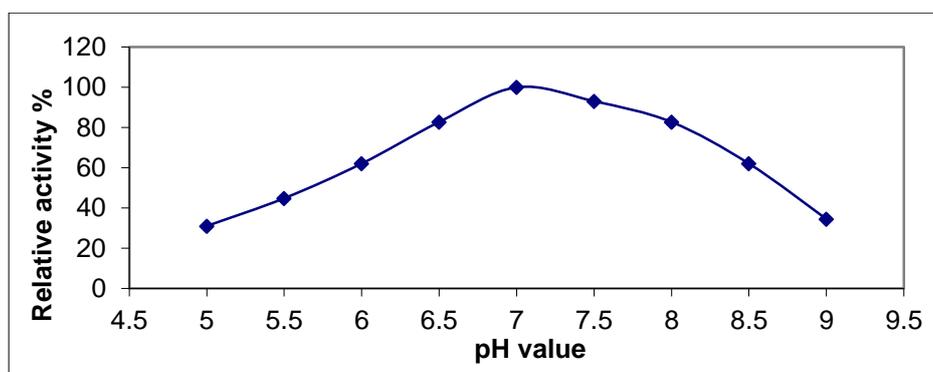


Fig (4): Effect of pH on the reaction activity of crude papaya pectinesterase (PE) enzyme.

Effect of temperature on the reaction activity of crude papain and papaya pectinesterase (PE) enzymes:-

Eleven different temperatures, i.e. 30, 35, 40, 45, 50, 55, 60, 65, 70, 75 and 80°C were chosen to investigate the optimum of crude papain enzyme. The experiments were carried out at optimum pH and the standard assay conditions. The obtained results are tabulated in Table (5) and illustrated in Fig. (5).

Maximum activity of crude papain was obtained at 60°C. The maximum rate of reaction for crude papain enzyme was found to be 8.64units/ml. Such values for optimum temperature and maximum activity are different with these previous study in which noted the optimal temperature of papain were 65°C, **Kilincet al. (2001)**,and 70°C,**Roy et al. (2005)**.

Table5.Effect of temperature on the reaction activity of crude papain enzyme.

Temperature (°C)	Obtained of tyrosine equivalents (µm/ml)	Activity (units/ml)	Specific activity U/mg protein	Relative activity (%)
30	0.38	4.56	6.16	52.77
35	0.52	6.24	8.43	72.22
40	0.54	6.48	8.75	75.00
45	0.49	5.88	7.94	68.05
50	0.51	6.12	8.27	73.44
55	0.69	8.28	11.19	95.83
60	0.72	8.64	11.68	100.00
65	0.66	7.92	10.70	95.04
70	0.65	7.80	10.54	93.60
75	0.60	7.20	9.72	86.40
80	0.33	4.00	5.41	46.20

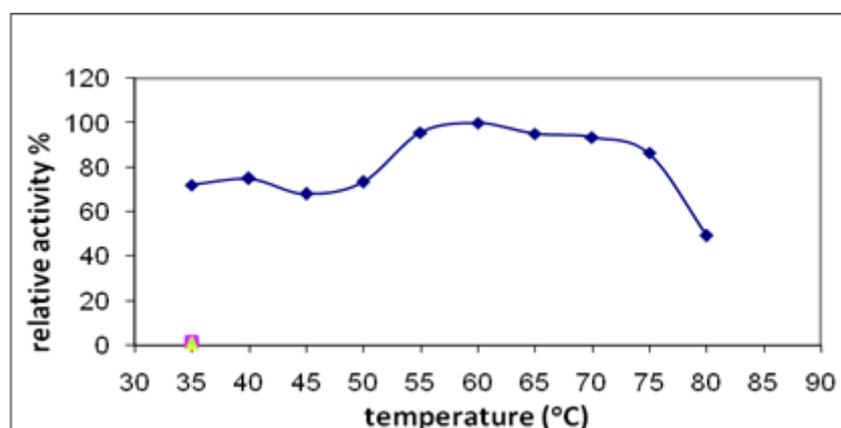


Fig (5): Effect of temperature on the reaction activity of crude papain enzyme.

Table 6. Effect of temperature on the reaction activity of crude papaya pectinesterase (PE) enzyme :-

Temperature (°C)	Activity (units/ml)	Specific activity U/mg protein	Relative activity (%)
30	9.0	18.75	45.50
35	10.8	22.50	54.50
40	13.2	27.50	66.60
45	15.6	32.50	78.80
50	17.4	36.25	87.90
55	19.8	41.25	100.00
60	18.6	38.75	93.90
65	18.0	37.50	90.91
70	16.8	35.00	84.85
75	12.0	25.00	60.61
80	10.2	21.25	51.52

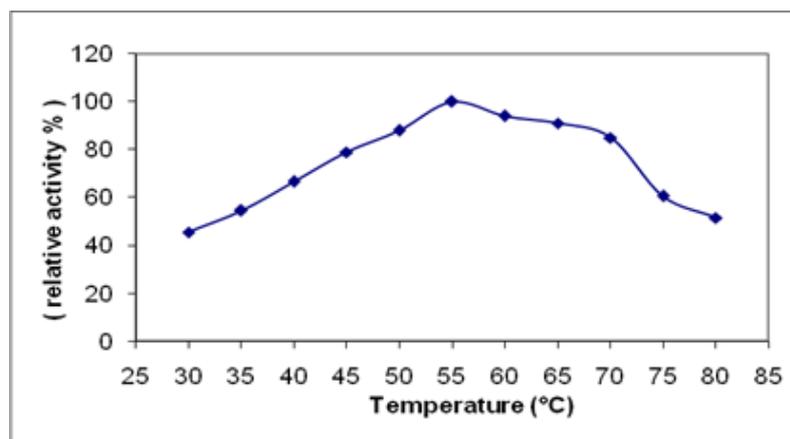


Fig (6): Effect of temperature on the reaction activity of crude papaya pectinesterase (PE) enzyme.

However, Papaya pectinesterase showed an optimum reaction temperature at 55°C Table (6) and illustrated in Fig (6). Over the temperature range (30- 80°C) was studied, the rate of deesterification was increased from 30° to 55°C until reached its maximum at 55°C and decreased after 70°C. The obtained optimum temperature was typical as found by **Maria *et al.* (2007)** who found 55°C to be the optimum temperature for pectinesterase from Hawthorn (*crataegus pubescens*) fruit. Also, it was close to those reported for the PE from apple by **Denes *et al.* (2000)**; bramley apple PE **King (1990)**; orange PE by **Korner *et al.* (1980)** and potato PE **Saenz *et al.* (2000)**. Those authors reported that 60°C was the optimum temperature for pectinesterase activities.

Effect of enzyme concentration on the reaction activity of crude papain and papaya pectinesterase (PE) enzymes:-

The effect of crude papain enzyme concentrations on the rate of enzymic hydrolysis of casein are presented in Table (7) and illustrated in Fig. (7). Ten different enzyme concentrations i.e. 0.1, 0.2, 0.3, 0.4, 0.5, 0.6, 0.7, 0.8, 0.9 and 1.0% (w/v) were tested for the optimal enzyme concentration at pH 6.4, 60°C and the incubation time was 10 min. The obtained results showed that the maximum activity reached to

9.6Units/ml at enzyme concentration of 0.6% (v/v). On the other hand, the reaction activity was decreased by increasing the amount of enzyme added. These light decreases might be attributed to the rate of the reverse reaction or inhibition effect which might be processed in opposite direction and steric hindrance is originated from the excess amount of enzyme. The obtained results are in agreement with that obtained by **Kilincet *et al.* (2001)** and **Roy *et al.* (2005)**.

While, the effect of different enzyme concentrations on the activity of crude papaya pectinesterase was evaluated. Different enzyme amounts i.e. 1.0, 2.0, 3.0, 4.0, 5.0, 6.0, 7.0, 8.0, 9.0, 10.0, 11.0, 12.0, 13.0, 14.0 and 15.0 ml were tested. The reaction activity of PE increased as the volume of enzyme concentration used increased till the volume of 12 ml as shown in Table (8) and illustrated in Fig (8). Then there was no further increment of activity as more enzymes were used. These observations may be attributed to the rate of the reverse reaction (inhibition effect) of the product which might produced in opposite direction leading to general decrease in the final product, **Illanes *et al.* (1998)**.

Table 7.Effect of enzyme concentration on the reaction activity of crude papain enzyme.

Enzyme conc. (%)	Obtained of tyrosine equivalents (µm/ml)	Activity (units/ml)	Specific activity U/mg protein	Relative activity (%)
0.1	0.52	6.24	8.43	65.00
0.2	0.60	7.2	9.72	75.00
0.3	0.64	7.68	10.37	80.00
0.4	0.65	8.2	11.08	85.41
0.5	0.70	8.40	11.35	87.5
0.6	0.80	9.60	12.97	100.00
0.7	0.74	8.88	12.00	92.50
0.8	0.73	8.76	11.83	91.25
0.9	0.70	8.40	11.35	87.50
1.0	0.70	8.40	11.35	87.50

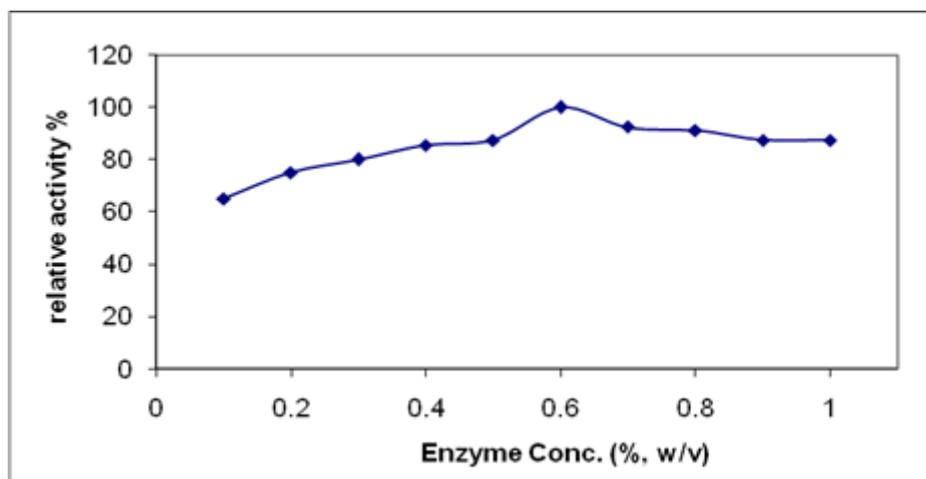


Fig (7): Effect of enzyme concentrations on the reaction activity of crude papainenzyme.

Table 8 .Effect of enzyme concentration on the reaction activity of crude papaya pectinesterase (PE) enzyme:-

Enzyme conc. (ml)	Activity (units/ml)	Specific activity U/mg protein	Relative activity (%)
1	1.2	2.5	8.82
2	2	4.16	14.68
3	2.8	5.83	20.57
4	4	8.33	29.40
5	5.2	10.83	38.23
6	6.4	13.33	47.05
7	7.6	15.83	55.88
8	9.2	19.16	67.63
9	10	20.83	73.53
10	11.2	23.33	82.35
11	12.4	25.83	91.18
12	13.6	28.33	100.00
13	13.6	28.33	100.00
14	13.2	27.5	97.07
15	13.2	27.5	97.07

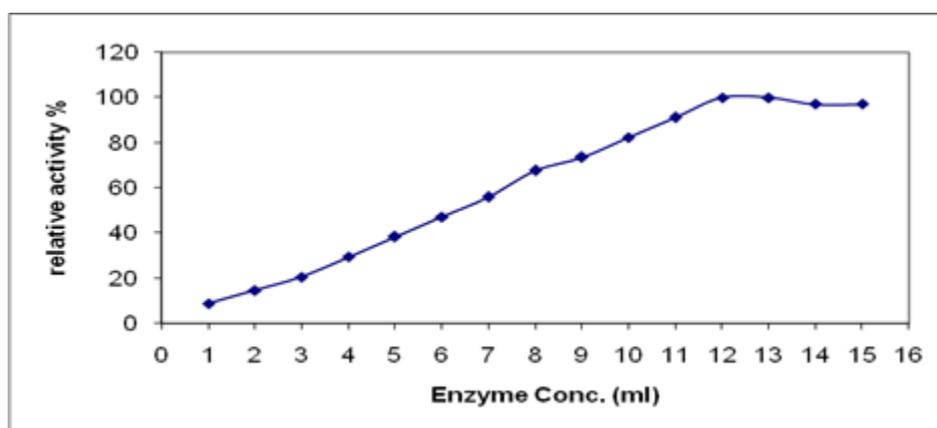


Fig (8): Effect of enzyme concentrations on the reaction activity of crude papaya pectinesterase (PE) enzyme.

Kinetic parameters of crude papain and papaya pectinesterase (PE) enzymes:-

The effect of substrate concentrations on the reaction velocity of crude papain enzyme was evaluated. The reaction mixture was carried out at different concentrations, i.e. (0.1, 0.2, 0.3, 0.4, 0.5, 0.6, 0.7, 0.8, 0.9, 1, 1.1, 1.2, 1.3, 1.4 and 1.5% (w/v) of casein as a substrate. The obtained results are illustrated in Table (9) and Fig. (9a, b). By plotting the obtained activity against the concentration of the substrate, the curve was obtained, Fig. (9a). From these results, the Michaelis-Menten constant and maximal reaction rate of the crude papain were determined and equalled to $K_m = 0.43$ g/100 ml phosphate buffer solution and $V_{max} = 13.68$ units/ml /min, respectively. The acquired K_m by **Lineweaver and Burk** plots (Fig 9c), was equally to that obtained by experimentally curve. These values are higher than that reported by **Shaw et al. (1987)** who found that kinetic parameters were calculated as 0.12 g/100 ml for K_m and 8.90 units/ml/min for V_{max} in the case of soluble papain enzyme. However, the activity of the crude papaya pectinesterase as a function of pectin concentration was also investigated. The reaction was carried out at different concentrations, i.e. 0.5, 1.0, 1.5, 2.0, 2.5, 3.0, 3.5, 4.0, 4.5, 5.0, 5.5, 6.0, 6.5, 7.0, 7.5, 8.0, 8.5, 9.0, 9.5 and 10.0 mg/ml of pectin as

a substrate. The obtained results were illustrated in Table (10) and Fig (10a,b). From these results, the Michaelis-Menten constant and maximal reaction rate of the crude papaya pectinesterase were determined and equalled to $K_m = 3.2$ mg/ml buffer solution and $V_{max} = 15.6$ units/ml, respectively.

The obtained K_m by **Lineweaver and Burk** plots (Fig. (10c) was equally to that obtained by experimentally curve. These values are different from that reported by **Maria et al. (2007)** who found that kinetic parameters were calculated as 2.84 mg/ml for K_m and 64.10 units/mg proteins for V_{max} in the case of pectinesterase enzyme from hawthorn fruits.

The present variation between the obtained K_m and V_{max} results of papain and pectinesterase may be due to that there were different concentrations of the both enzymes in one ml of the different experiments which related to the different sources of papain or pectinesterase extracted or obtained.

From the obtained results, it can conclude that papaya fruits contain benefits industrial enzymes such as papain and pectinesterase. Therefore the research is concentrated on the optimum conditions (PH, temperature, enzyme concentration, substrate concentration) about the production and the activity of both enzymes to achieve ideal uses of them.

Table 9. Effect of substrate concentrations on the reaction velocity of crude papain enzyme.

Substrate concentration (%)	[1/S]	Activity Units/ml	Reaction velocity (V)*	[1/V] ×10 ⁻¹
0.1	10.00	0.96	2.58	3.87
0.2	5.00	3.00	4.34	2.30
0.3	3.33	4.32	5.62	1.78
0.4	2.50	6.72	6.59	1.52
0.5	2.00	8.04	7.35	1.36
0.6	1.66	8.40	7.96	1.26
0.7	1.43	10.32	8.47	1.18
0.8	1.25	11.88	8.89	1.12
0.9	1.11	12.96	9.26	1.07
1.0	1.00	13.56	9.56	1.04
1.1	0.91	13.68	9.84	1.01
1.2	0.83	12.36	10.07	0.99
1.3	0.77	12.12	10.28	0.97
1.4	0.71	11.64	10.47	0.95
1.5	0.66	11.52	10.63	0.94

 $V_{max} = 13.68$ Units/ml. $K_m = 0.43$ g/100 ml

$$* V = \frac{V_{max} \times [S]}{K_m + [S]}$$

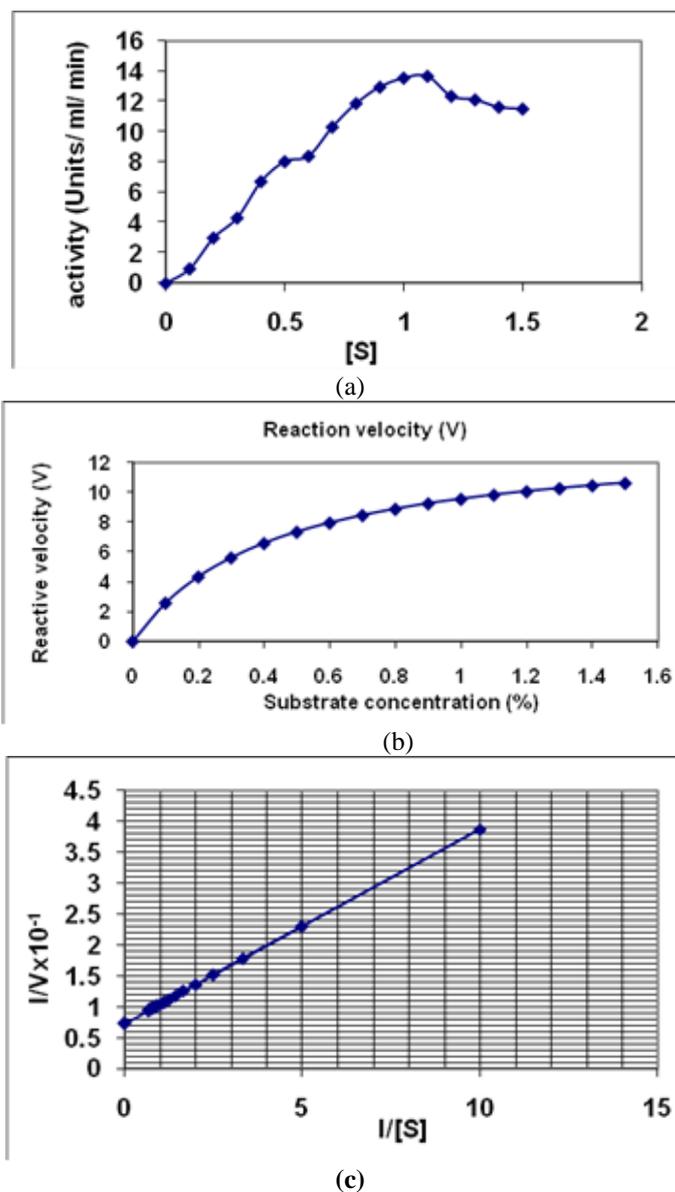


Fig (9): Effect of substrate concentrations on the reaction velocity of crude papain enzyme.

Table 10. Effect of substrate concentrations on the reaction velocity of crude papaya pectinesterase (PE) enzyme.

Substrate concentration (mg/ml)	$1/[S]$	Activity Units/ml	Reaction velocity (V)*	$[1/V] \times 10^{-1}$
0.5	2.00	2.7	2.11	4.73
1.0	1.00	3.6	3.71	2.69
1.5	0.66	4.5	4.98	2.00
2.0	0.50	5.7	6.00	1.66
2.5	0.40	6.3	6.84	1.46
3.0	0.33	7.2	7.55	1.32
3.5	0.28	9.0	8.15	1.22
4.0	0.25	9.6	8.66	1.15
4.5	0.22	10.8	9.12	1.09
5.0	0.20	11.7	9.51	1.05
5.5	0.18	12.6	9.86	1.01
6.0	0.166	14.7	10.17	0.98
6.5	0.153	15.6	10.45	0.95
7.0	0.142	13.5	10.71	0.93
7.5	0.133	12.6	10.93	0.91
8.0	0.125	12.0	11.14	0.89

8.5	0.117	11.7	11.33	0.88
9.0	0.111	10.5	11.51	0.86
9.5	0.105	10.5	11.66	0.85
10	0.100	9.9	11.82	0.84

$V_{max} = 15.6 \text{ Units/ml.}$ $K_m = 3.2 \text{ mg/ml}$ * $V = \frac{V_{max} \times [S]}{K_m + [S]}$

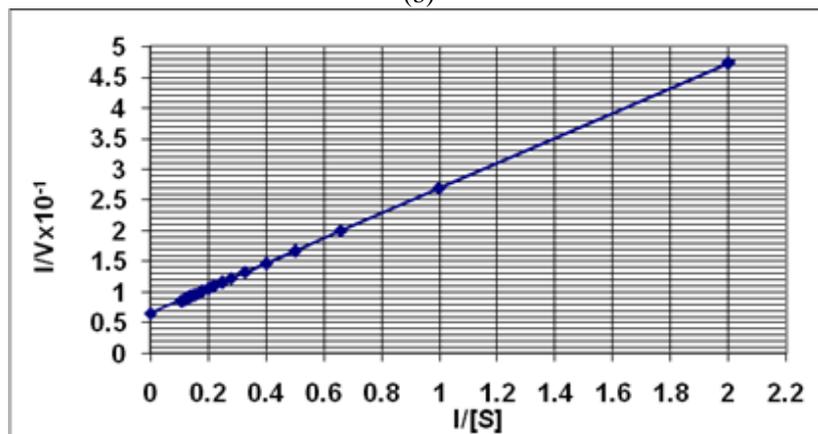
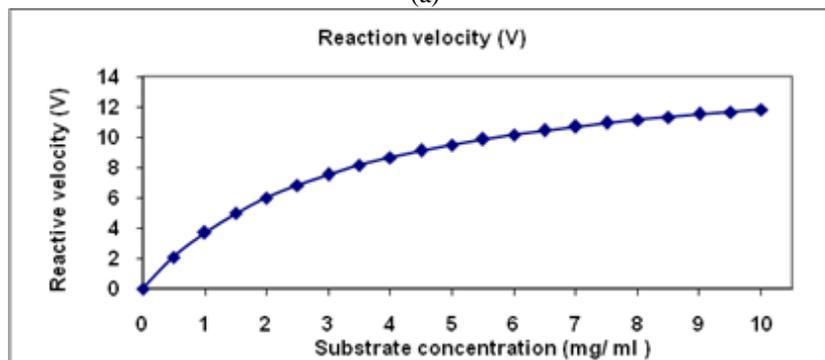
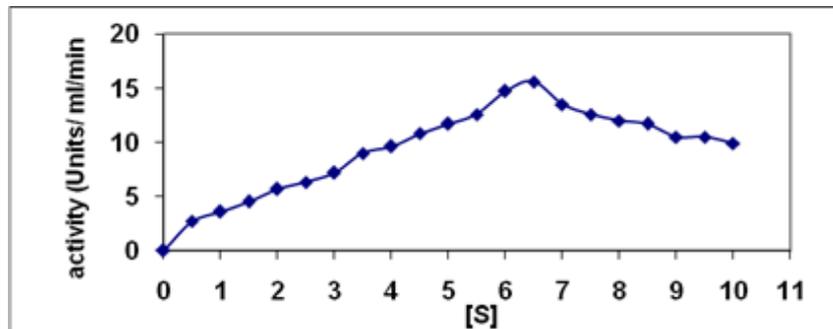


Fig. (10): Effect of substrate concentrations on the reaction velocity of crude papaya pectinesterase (PE) enzyme.

References

Arnon, R. (1970). Papain. Proteolytic enzymes. In: Pearlman GE, editor. Methods of enzymology, vol. XIX. New York: American press; P.226.

Baeza, G.; Correa, D. and Salas, C. (1989). Proteolytic enzymes in *Caricacandamarcensis* J. Sci. Food. Agric., 51:1-9.

Baron, A. and Thibault, J.F. (1985), Les enzymes pectolytiques. In: Hydrolases etDépolimérasés, ed. A. Mouranche&C.Costes. Gauthier-Villars, Paris, pp. 143-164.

Bradford, M.M. (1976). A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein dye binding. Biochem, 72: 248-254.

- Dayanand, C.D. (2013).** Evaluation of comparative total proteolytic activity in plant lattices. *Int. J. Life sci. Bt Pharm. Res.*, 2(1): 47-55.
- Denes, J.M.; Baron, A. and Drillcau, J.F. (2000).** Purification properties and heat inactivation of pectinmethyl-esterase from apple. *J. Sci. Food Agric.* 80 (10): 1503-1509.
- Fayyaz, A. and Asbi, B.A. (1993).** Pectinesterase extraction from guava. *Pertanika J. Trop. Agric. Sci.* 16 (3): 223-224.
- Fayyaz, A.; Asbi, B. A.; Ghazali, H. M.; Che Man, Y. B. and Jinap, S. (1995).** Kinetics of papaya pectinesterase. *Food Chem.* 53: 129-135.
- Illanes, A.; Altamirano, C.; Aillapan, A.; Tomasello, G. and Zuniga, M. (1998).** Packed-bed reactor performance with immobilized lactase under thermal inactivation. *Enzyme and Microbial Technol.*, 23(1-2): 3-9.
- Teixeira da Silva, JA.; Zinia, R.; Nhut, D.T.; Sivakumar, D.; Gera, A.; Souza Jr, M.T. and Tennant, P. (2007):** papaya (*Carica papaya* L.) biology and biotechnology. *Tree and forestry science and biotechnology.* 1(1), 47-73.
- Kertesz, Z. (1955).** Pectic enzymes. In: *Methods in enzymology*, S.P. Colowick and N.O. Kaplan. Academic press, New York, USA. 1:158.
- Kilinc, A.; Onal, S. and Telefoncu, A. (2001).** Stabilization of papain by modification with chitosan. *Turk. J. Chem.* 26: 311-316.
- King, K. (1990).** Partial Characterization of the in situ activity of pectinesterase in Bramely apple. *Int. J. Food Sci. Technol.* 25: 188-197.
- Korner, B.; Zimmermann, G. and Berk, Z. (1980).** Orange pectinesterase: Purification properties and effect on cloud stability. *J. Food Sci.* 45: 1203-1206.
- Lineweaver, H. and Burk, D. (1954).** Methods of plotting enzyme kinetic data. *J. Am. Chem. Soc.*, 56: 658-663.
- Maria, A. V.; Juan, A. S.; Graciano, C. and Emma, G. R. (2007).** Extraction, thermal stability and kinetic behavior of pectinmethyl-esterase from hawthorn (*Crataegus pubescens*) fruit. *LWT* 40: 278-284.
- Nikolić, M.V. and Mojovic, L. (2007).** Hydrolysis of apple pectin by the coordinated activity of pectic enzymes. *Food Chemistry*, 101: 1-9.
- Nitsawang, S.; Hatti-Kaul, R. and Kanasawuda (2006).** *Enzyme Microb. Technol.*, 39:1103-1107.
- Pamela R, James M. (2010):** Genetically Engineered Distortions. *The New York Times*, May 14. Retrieved August 10, 2010.
- Papain and other constituents of *Carica papaya* L. *Top. Enzyme Ferment. Biotechnol.*, v. 5, p. 262 – 35.
- Pitprecha, S. and Damrongsakkul, S. (2006).** Hydrolysis of raw hide using proteolytic enzyme extracted from papaya latex. *Korean J. Chem. Eng.*, 23(6): 972-976.
- Polacsek-Ra'cz, M. and Posar-Hajnal, K. (1976).** Determination of pectinmethyl-esterase, polygalacturonase and pectic substances in some fruits and vegetables. Part II. Study into the pectolytic tissue enzymes of Jonathan apples and some other fruits and vegetables. *Acta Alimentaria*, 5: 189-204.
- Puig, A.; Gil, I. and Sanchez, O. (2008).** Evaluation of drying techniques measuring proteolytic activity of papain obtained from unripe fruit and skin juice. *Int. conf. industrial biotechnol.*, 14: 345-350.
- Rao, P. S. and Rangan, R. K. (2001).** Production of crude papain from papaya latex and by-product utilization of papaya. *Horticultural J.*, 14(3):1-5.
- Rofael, w. and Salwa, D. (2003).** Preparation of crude and refined papain from papaya latex. *J. Agric. Sci. Mansoura univ.*, 28(5): 3783-3790.
- Roy, J.J.; Sumi, S.; Sangeetha, K. and Abraham, T.E. (2005).** Chemical modification and immobilization of papain. *J. Chem. Technol. Biotechnol.* 80: 184-188.
- Saenz, J.M.; Tellez, A.; Garza, H.; Reyes, M.; Contreas Esquivel, J.C.; Aguilar, C.N. and Garza, H. (2000).** Purification and some properties of pectinesterase from potato. *Brazilian Archives of Biology and Technology* 43(4): 393-398.
- Sarr, F. and Tsai Pi Jen, (2008).** Effects of acidification on PE activity, color and antioxidant properties of cold break tomato juice. *J. Food Quality*, 31(1): 34-47.
- Shaw, J.; Change, R. and Wang, Y. (1987).** Kinetics of papain immobilized on chitosan by multiple point attachment. *Bot. Bull. Academic sinica*, 28: 131-138.
- Van den broeck, I.; Ludikhuyze, L.; Weemaes, C.; Van loey, A. and Hendrickx, M. (1999).** Thermal inactivation kinetics of pectinesterase extracted from oranges. *J. Food Processing and Preservation*, 23(5):391-406.
- Wicker, L. (1992).** Selective extraction of thermostable pectinesterase. *J. Food Sci.*, 57: 534-535.

إنتاج وتقييم إنزيم البابين والبكتين إستيريز من ثمار الباباظ

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تهدف هذه الدراسة الى إنتاج بعض الأنزيمات مثل انزيم البابين وانزيم البكتين استيريز من ثمار الباباظ ودراسة افضل الظروف للحصول على اعلى معدل استخلاص لهم.

دراسة العوامل التي تؤثر على سرعة التفاعل والنشاط لكلا من انزيم البابين والبكتين استيريز من تركيز القوة الأيونية ل NaCl , PH , درجة الحرارة , تركيز الأنزيم, تركيز مادة التفاعل المستخدمة وإيجاد كلا من ثابت ميكاليس K_m والسرعة القصوى V_{max} لامكانية التوصل الى انسب الظروف المثلي لنشاط هذه الانزيمات.

اوضحت النتائج ان كمية العصارة اللبنية المستخلصة كانت 6 جم /كجم ثمار وبعد التجفيف كانت الكمية 1,5 جم /كجم ثمار حيث ان نسبة الرطوبة كانت 74,6% للعصارة اللبنية .

وجد ان افضل تركيز من كلوريد الصوديوم لاستخلاص انزيم البكتين استيريز كانت 1,5 مولر حيث وصل نشاط الإنزيم الى اعلى معدل لنشاط وهو 6,25 وحدة/ ملجم بروتين.

كذلك وجد ان درجة النشاط والمحتوى البروتيني لانزيم البابين هي 7,68 وحدة /ملل و 0,74 ملجم/ ملل على التوالي. كذلك بالنسبة لانزيم البكتين استيريز كانت 2,8 وحدة /ملل و 0,48 ملجم / ملل على التوالي.

وجد ان افضل تركيز من كلوريد الصوديوم المطلوب لنشاط انزيم البكتين استيريز هو 0,3 مولر .

ان افضل درجة pH اللازم لنشاط كلا من البابين والبكتين استيريز هو 6,4 و 7 على التوالي .

ان افضل درجة حرارة اللازمة لنشاط كلا من البابين والبكتين استيريز هي 60 و 55 درجة مئوية على التوالي .

ان افضل تركيز لانزيم اللازم لنشاط كلا من البابين والبكتين استيريز هو 0,6 جم/100ملل محلول منظم و 12 مل على التوالي .

ان افضل تركيز للمادة المتفاعلة اللازمة لنشاط كلا من البابين والبكتين استيريز هو 1,1 جم/100ملل محلول منظم و 6,5 ملجم/ملل.

كذلك وجد ان قيمة كلا من V_{max} , K_m لكلا من انزيم البابين والبكتين استيريز هي 0,43 جم / 100مل محلول منظم , 13,68

وحدة/ ملل بالنسبة للبابين و 3,2 جم/ 100ملل و 15,6 وحدة/ ملل بالنسبة للبكتين استيريز على التوالي .